## GROSS PATHOLOGICAL FINDINGS OF AFRICAN SWINE FEVER SUSPECTS IN OEBELO, KUPANG REGENCY, 2021

# Maxs U. E. Sanam<sup>1</sup>, Maria Aega Gelolodo<sup>1\*</sup>, Larry R. W. Toha<sup>1</sup>, Tri Utami<sup>2</sup>, Yohanes T. R. M. R. Simarmata<sup>2</sup>, Theresia F. I. M. D. Murni<sup>3</sup>

<sup>1</sup>Department of Veterinary Diseases and Veterinary Public Health, Faculty of Veterinary Medicine, Universitas Nusa Cendana, Kupang, NTT, Indonesia 85001

<sup>2</sup>Department of Clinical, Reproductive, Pathology, and Nutrition, Faculty of Veterinary Medicine, Universitas Nusa Cendana, Kupang, NTT, Indonesia 85001

<sup>3</sup>Animal Husbandry Department of Kupang Regency, Kupang Regency, NTT, Kabupaten Kupang, NTT, Indonesia \*Corresponding author: gelolodo.m@staf.undana.ac.id

#### ABSTRACT

African swine fever (ASF) is a destructive re-emerging swine disease that has posed a serious economic threat to the global pig farming sector. In past years, ASF has rapidly spread over Europe, Asia, and Oceania, and begin to enter Indonesia in the middle of 2019. The clinical and pathological symptoms of ASF are influenced by the strain's virulence, the transmission pathway, and the pig's immunological and health status. ASF's clinical manifestations are known to evolve, from after an invasion enters a new free region to after the disease has been established in the territory for a longer period. Identifying ASF clinical signs and pathological changes is crucial for a comprehensive and reliable early detection system. The objective of this research is to observe and identify gross pathology in ASF suspect pigs in order to obtain a better understanding of the cause of death. Two dead pigs from a farm in Oebelo village, Kupang regency, Indonesia with a recent history of massive deaths had been examined in this study. The post-mortem results showed that hemorrhagic splenomegaly and hemorrhagic lymphadenitis were the main lesions observed at the examinations. Furthermore, hemorrhages were also found in various internal organs such as the kidneys, liver, and heart. To determine the exact cause of the pigs' deaths, a molecular diagnostic test should be conducted.

Keywords: African swine fever, virus, pigs, gross pathology, Kupang

#### **INTRODUCTION**

African swine fever (ASF) is a highly infectious hemorrhagic disease that affects domestic pigs and wild boars globally, posing a significant threat to swine health and welfare (OIE, 2019). Due to the high level of morbidity and mortality, stamping-out policies, restrictions on traffic or trade-in pork or its products, and the absence of effective vaccines and treatment, this virus poses a serious threat to the swine industry and global socio-economy (Bosch-Camós et al., 2020; Costard et al., 2013). Although ASF is a nonzoonotic disease with no direct impact on public health, (Dixon et al., 2020: OIE. 2019). it threatens food security (Costard et al., 2013), particularly for backyard and smallholding pig farmers in developing countries who rely on pigs not only for additional income but also for the source of animal protein (Sanchez-Cordon et al., 2018).

ASF is caused by the African Swine Fever Virus (ASFV), a doublestranded DNA virus belonging to the Asfarviridea family and the genus Asfarvirus, which is the only DNA arbovirus (OIE, 2019). ASFV infects only Suidae family members of all ages, however, just domestic pigs and wild boars acquire the clinical disease. whereas warthogs *aethiopicus*) (Phacochoerus and bushpigs (Potamochoerus porcus) act as asymptomatic carriers and reservoirs of ASFV (Beltrán-Alcrudo et al., 2017). In terms of disease epidemiology, ASF is known to have a very complicated epidemiological pattern since it incorporates multiple different epidemiological cycles, such as the sylvatic cycle, the tick-pig cycle, the domestic cycle, and the wild boar-habitat cycle (Chenais et al., 2019; Viltrop et al., 2021). ASFV transmission is facilitated by several variables, including infected pigs, contaminated pork products, fomites,

and the *Ornithodoros* vector (Blome *et al.*, 2020; Gaudreault *et al.*, 2020).

The clinical form of ASF from peracute, ranges acute. subacute, chronic to asymptomatic, depending on the virus strain and virulence, transmission route, dosage, host features, and endemicity status in a region (Beltrán-Alcrudo et al., 2017; Pedro J. Sánchez-Cordón et al., 2020; P.J Sánchez-Cordón et al., 2021). There are 24 genotypes of ASFV that circulate strains worldwide, and they are classified as highly, moderately, or low virulent (Urbano et al., 2021). The gross pathological findings of this disease vary according to the disease's course and the diversity of the aforementioned clinical characteristics and contributing factors (Blome et al., 2020; Sánchez-Vizcaíno et al., 2015). Thus, OIE (2019) suggested that ASF diagnosis should be based on epidemiological observation studies, of clinical symptoms, gross pathology, and histopathological findings, and a series of laboratory tests (OIE, 2019).

ASFV has remained endemic in Africa since its first identification in Kenya in 1921, impacting up to 35 African nations. All 24 genotypes of ASFV are reported to be present in Africa. ASFV genotype II first appeared outside Africa in the Republic of Georgia in 2007, and since then it has expanded across the Caucasus area, as well as into the Russian Federation and Eastern Europe, where it has continued to circulate and spread to other parts of the world (Gaudreault *et al.*, 2020). Virus ASF was first detected in China, which has half of the world's pig population, in August 2018 (Zhou *et al.*, 2018), from China, ASFV is quickly spreading over Eur ope, Asia, and the Pacific (Brookes *et al.*, 2021). The rapid and extensive spread of this disease has boosted awareness of the threat caused by this serious disease not just to the global economy but also to food security (FAO, 2021).

ASF was detected for the first time in Indonesia around the end of 2019 in North Sumatra (Dharmayanti et al., 2021). As of September 2021, ASF cases had been recorded in ten provinces in Indonesia, including North Sumatra, Riau, West Sumatra, South Sumatra, Lampung, West Java, Central Java, Yogyakarta, Bali, and Nusa Tenggara Timur/NTT (from a total of 34 provinces in Indonesia) (FAO, 2021). According to Dharmayanti et al. (2021), the ASF virus that is responsible for the outbreaks in North Sumatra and West Java is ASFV Genotype II, which belongs to a cluster of Vietnam, and Georgia, China, Belgium. Information regarding the ASFV genotype circulating in other parts of Indonesia is still not available until now.

ASF infections in NTT were initially identified on Timor Island in early 2020 and have since spread to other regions of the province. Currently, 20 districts (including Kupang Regency) and 1 city in NTT have been reported to be infected with the ASF virus. Only one district in the region has yet to report an ASF occurrence. Strict supervision and monitoring are carried out in this area (OIE, 2021). Even today, there is limited information available on the situation and progress of ASF in NTT.

Even though ASF has been known for almost a century, attempts to prevent and manage it is proving to be a major challenge for global animal health. (Sanchez-Cordon et al., 2018). Early detection, strict surveillance. epidemiological investigation, culling of sick pigs, quarantine, biosecurity, and monitoring of the pigs and the movements of their products are the only solutions to protect or control the spread of ASF (Beltrán-Alcrudo et al., 2017). It is believed that the **ASF-infected** earlier pigs are identified, the quicker veterinary authorities can act to determine further diagnostic testing methods and the next measure in disease control. The purpose of this study is to examine the gross pathological features of ASF-suspected pigs in order to give a snapshot of ASF incidence in endemic areas. Thus, authorities are assisted in establishing a set of control policies for their territory and neighboring areas.

## MATERIALS AND METHODS

Macroscopic lesions were observed on two domestic pigs necropsied immediately after deaths on a farm in Oebelo Village, Kupang Regency, in March 2021.

Necropsies were performed on deceased pigs at the farm's burial

site. The biosecurity protocol according to FAO's A Manual for Veterinarians (Beltrán-Alcrudo *et al.*, 2017) was implemented on handling of carcasses, materials, tools, the personnel involved, and carcasses disposal management.

### **RESULTS AND DISCUSSION**

According to the farmer's information, the farm was currently experiencing a large number of pig deaths. All of the pigs on the farm were reportedly dead in less than a month, with similar clinical manifestations. All the pigs on the farm had already been vaccinated against CSF (*Classical Swine Fever*). The farmer recorded that prior to death, the pigs displayed symptoms in

the form of fever ranging from 41-42° C, weakness, laziness, crowding in one spot, and loss of appetite. Pigs on the farm mostly die less than a week after exhibiting clinical symptoms. Hemorrhages were discovered in various areas of the pig's skin, notably around the eyes, ears, abdomen, caudal skin area, and legs, according to post-mortem observations (Figure 1).



Figure 1. (A) Petechiae and ecchymoses around eyes and facial area; (B) Ecchymoses, a hematoma on ears; (C) Petechiae in the abdomen; and (D) Petechiae in the caudal skin area.

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Furthermore, the results of postmortem examination showed hemorrhagic lesions in multiple organs of the pig. The spleens appear darker and enlarged (splenomegaly), hemorrhage and enlargement were also observed in the lymph nodes, petechial and ecchymoses in the renal cortexes, hearts with interstitial hemorrhages, and hepatomegaly with hemorrhages (Figure 2). Previously, an investigation into a case of mass death in pigs on another farm in the same area also showed similar clinical and pathology finding in the

form of multiorgan hemorrhage (Gelolodo et al., 2021). Both of these farms' pigs had been vaccinated with CSF. This raises the suspicion of an outbreak caused by а highly contagious infectious disease. These two farms are located in an area known to be endemic to CSF and ASF. ASF and CSF are two major pig diseases with similar clinical and pathological signs characterized by immunosuppression and hemorrhage. diseases Those can result in significant mortality rates in pigs (Schulz et al., 2017).

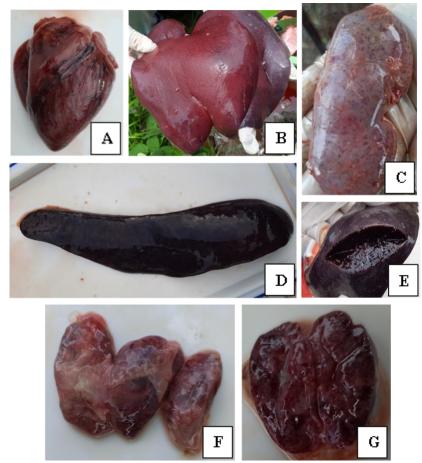


Figure 2. (A) Heart with interstitial hemorrhages; (B) Hepatomegaly and unevenly colored liver with subcapsular hemorrhage (C) Diffuse petechiae and ecchymoses in the renal cortex; (D&E) Spleen with an increase in size, rounded edges, and dark red to black color (hyperaemic splenomegaly); and (F&G) enlarged, edematous and hemorrhagic lymph nodes.

As previously stated, the clinical presentation and gross pathological manifestations of ASF in domestic pigs might differ based on the viral isolate's virulence, the method and dose of infection, and the host's features (Salguero, 2020; Sánchez-Vizcaíno et al., 2015). The diversity of clinical manifestations is also matched by the diversity of immunological responses, implying a web complex of virus-host interactions (P.J Sánchez-Cordón et al., 2021). Regarding the virulence factor, high virulence ASFV typically induces per acute and acute infection, ASFV with medium virulence cause acute and subacute infection, whilst ASF viruses with low virulence generate moderate and nonspecific symptoms that are easily misled with other diseases (Beltrán-Alcrudo et al., 2017; P.J Sánchez-Cordón et al., 2021).

The per acute type of ASF is marked by a brief incubation period, high fever (41-42 °C), lack of appetite, and laziness. Unexpectedly death can occur within 1-3 days before the appearance of any clinical symptom (Beltrán-Alcrudo et al., 2017). Pigs infected with this form typically die without showing any macroscopic exterior or internal pathologies and only show nonspecific symptoms (Sánchez-Vizcaíno et al., 2015).

The acute infection is the most prevalent ASF clinical manifestation in natural outbreaks (P.J Sánchez-Cordón *et al.*, 2021) and the most common clinical form seen in naive pigs (Salguero, 2020). Under natural conditions, the incubation time for acute ASF ranges from 4 to 19 days al.. (Sanchez-Cordon et 2018: Sánchez-Vizcaíno et al., 2015) with death occurring within 4-10 days. This form's case-fatality rate can be as high as 100% (OIE, 2019). A febrile state (40.5-42 °C) with erythema and cyanosis of the skin generally characterizes this type (Gallardo et al., 2015). Erythema appears on affected pigs' ears, chest, abdomen, distal extremities, tails, and perianal area. Cyanotic patches generally form in these places around 24-48 before hours death. Ecchymoses and petechial hemorrhages can also be seen on the skin. Pigs in the same cage also huddle or pile on top of one another, displaying loss of appetite, inactivity, frailty, recumbency, and an inability to stand up (P.J Sánchez-Cordón et al., 2021; Sánchez-Vizcaíno et al., 2015). Internal lesions such as hemorrhagic splenomegaly, hemorrhagic lymphadenitis, and renal petechiae are the most common in this form (Gallardo et al., 2015; Salguero, 2020). The affected spleen is usually enlarged with a rounded margin, friable when sectioned, and dark in color (Sánchez-Vizcaíno et al., 2015). Lymph nodes, primarily the gastrohepatic, renal (Sánchez-Vizcaíno et al., 2015), as well as other abdominal lymph nodes such as ileocecal and mesenteric exhibit medulla hemorrhages, hence why areas of damaged lymph nodes have a marbled pattern. Other lymph nodes,

like the submandibular. retropharyngeal, or inguinal, may also have hemorrhages with a lower frequency (Salguero, 2020). Petechial hemorrhages in the cortex and renal pelvis are common in kidneys. Other lesions seen in pigs with acute versions of the disease include petechial hemorrhages in the urinary bladder mucosa, epicardium, endocardium, and pleura (Salguero, 2020; Sánchez-Vizcaíno et al., 2015).

Furthermore, in the subacute type of ASF, death usually occurs within 7-20 days (Beltrán-Alcrudo et al., 2017; Galindo-Cardiel et al., 2013), and the mortality rate might range between 30% and 70%. Survivors of this form usually recover after 3-4 weeks (Sánchez-Vizcaíno et al., 2015). In this form, some pigs may not show any clinical indications, while others may show very minor signs (P.J Sánchez-Cordón et al., 2021). The clinical signs are similar to those seen in animals with the acute form of the disease, however, the indications clinical are less pronounced (Sánchez-Vizcaíno et al., 2015) and can lasting longer until 4 weeks (P.J Sánchez-Cordón et al., 2021). However, the vascular abnormalities are shown in subacute forms of ASF, which are marked by widespread hemorrhages in the lymph nodes, kidneys, and spleen, as well as diffuse organ enlargement, are more severe than those seen in the acute type (Galindo-Cardiel et al., 2013; Gómez-Villamandos et al., 2013). The occurrence of subacute and

chronic forms is less common than the acute form (Salguero, 2020).

The findings of this case study suggest that the clinical signs and pathological gross findings are similar to acute and subacute forms of ASF, which are characterized by multiple visceral organ hemorrhages, particularly in the spleen, lymph nodes, and kidney (Salguero, 2020; Yamada et al., 2021). According to C. Gallardo et al. (2021), ASFV genotype I and II are known as the most widespread ASF genotype in the world and responsible for many acute and subacute cases in many endemic In Asia, particularly areas. in Indonesia, genotype II is currently the most common ASFV genotype (Dharmayanti et al., 2021). An experiment using a highly virulent ASFV genotype II strain revealed that the genotype's gross findings were consistent with acute ASFV infection (Rodríguez-Bertos et al., 2020). Izzati et al. (2020) also found that genotype II can cause ASF in both acute and subacute forms. This study showed that the acute forms of ASFV genotype II from the Vietnam isolate displays hemorrhagic lesions in multiple organs, including hyperemic splenomegaly, hemorrhagic lymph nodes, petechial hemorrhage in the renal cortex, and hemorrhages in the epicardium, while the subacute form showed more severe hemorrhagic and edema toward the visceral organs (Izzati et al., 2020). Nga et al. (2020) also reported a similar result, indicating that acute forms caused by ASFV genotype II exhibited the

same gross pathology features as previously mentioned. Other studies about acute forms caused by ASF genotype II from Rusia (Kolbasov *et al.*, 2018), China (Zhao *et al.*, 2019), Estonia, and Poland (Carmina Gallardo *et al.*, 2021) isolates also reported similar gross pathology characteristics. Nonetheless, other aspects of diagnosis, such as laboratory diagnosis, particularly molecular diagnostic as which is the gold standard of ASF diagnosis, should be utilized to verify the case in order to obtain a comprehensive and definitive diagnosis of ASF.

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### **CONFLICT OF INTEREST**

The authors confirm that there is no conflict of interest in this study.

### REFERENCES

- Beltrán-Alcrudo, D., Arias, M., Gallardo, C., Kramer, S. A., & Penrith, M.-L. (Eds.). (2017).
  African swine fever: detection and diagnosis – A manual for veterinarians (FAO Animal Production and Health Manual No. 19 ed.): Food and Agriculture Organization of the United Nations (FAO).
- Blome, S., Franzke, K., & Beer, M. (2020). African swine fever -A review of current knowledge. *Virus Res*, 287, 198099. doi:10.1016/j.virusres.2020.1 98099
- Bosch-Camós, L., López, E., & Rodriguez, F. (2020). African swine fever vaccines: a

promising work still in progress. *Porcine Health Manag*, 6, 17. doi:10.1186/s40813-020-00154-2

- Brookes, V. J., Barrett, T. E., Ward, M. P., Roby, J. A., Hernandez-Jover, M., Cross, E. M., . . . Khalfan, S. (2021). A scoping review of African swine fever virus spread between domestic and freeliving Transbound pigs. Emerg Dis, 68(5), 2643-2656. doi:10.1111/tbed.13993
- Chenais, E., Depner, K., Guberti, V., Dietze, K., Viltrop, A., & Ståhl, K. (2019). Epidemiological considerations on African

swine fever in Europe 2014– 2018. *Porcine Health Management*, 5(1), 6. doi:10.1186/s40813-018-0109-2

- Costard, S., Mur, L., Lubroth, J., Sanchez-Vizcaino, J. M., & Pfeiffer, D. U. (2013). Epidemiology of African swine fever virus. *Virus Research*, *173*(1), 191-197. doi:https://doi.org/10.1016/j.v irusres.2012.10.030
- Dharmayanti, N. I., Sendow, I., Ratnawati, A., Settypalli, T.
  B. K., Saepulloh, M., Dundon,
  W. G., . . . Lamien, C. E.
  (2021). African swine fever in North Sumatra and West Java provinces in 2019 and 2020, Indonesia. *Transbound Emerg Dis.* doi:10.1111/tbed.14070
- Dixon, L. K., Stahl, K., Jori, F., Vial, L., & Pfeiffer, D. U. (2020). African Swine Fever Epidemiology and Control. *Annual Review of Animal Biosciences*, 8(1), 221-246. doi:10.1146/annurev-animal-021419-083741
- FAO. (2021). ASF situation in Asia and Pacific Update per 2 September 2021. Retrieved from http://www.fao.org/ag/againf o/programmes/en/empres/AS F/situation\_update.html.
- Galindo-Cardiel, I., Ballester, M., Solanes, D., Nofrarías, M., López-Soria, S., Argilaguet, J. M., . . . Segalés, J. (2013). Standardization of

pathological investigations in the framework of experimental ASFV infections. *Virus Research, 173*(1), 180-190. doi:https://doi.org/10.1016/j.v irusres.2012.12.018

- Gallardo, C., Sastre, P., Rueda, P., Gerilovych, A. P., Kit, M., Nurmoja, I., & Potier, M. F. L. (2021). 5. Methods for African swine fever diagnosis in clinical and environmental samples. In Understanding and combatting African Swine Fever (pp. 141-160): Wageningen Academic Publishers.
- Gallardo, C., Soler, A., Nurmoja, I., Cano-Gómez, C., Cvetkova, S., Frant, M., . . . Nieto, R. (2021). Dynamics of African swine fever virus (ASFV) infection in domestic pigs infected with virulent, moderate virulent, and attenuated genotype II ASFV European isolates. Transboundary and Emerging Diseases.
- Gallardo, M. C., Reoyo, A. d. l. T., Fernández-Pinero, J., Iglesias, I., Muñoz, M. J., & Arias, M. L. (2015). African swine fever: a global view of the current challenge. *Porcine Health Management, 1*(1), 21. doi:10.1186/s40813-015-0013-y
- Gaudreault, N. N., Madden, D. W., Wilson, W. C., Trujillo, J. D., & Richt, J. A. (2020). African

Jurnal Kajian Veteriner

SwineFeverVirus:AnEmergingDNAArbovirus.Frontiersinveterinaryscience,7(215).doi:10.3389/fvets.2020.00215

- Gelolodo, M. A., Sanam, M. U., Toha, L. R., Widi, A. Y., Simarmata, Y. T., & Murni, T. (2021). Histopatologi F. Limpa dan Limfonodus pada Kasus Lapangan dengan Dugaan Kematian Akibat Virus African Swine Fever Pada Babi di Kabupaten Kupang. Jurnal Kajian Veteriner, 9(2), 62-75.
- Gómez-Villamandos, J. C., Bautista, M. J., Sánchez-Cordón, P. J., & Carrasco, L. (2013).
  Pathology of African swine fever: The role of monocytemacrophage. *Virus Research*, *173*(1), 140-149. doi:https://doi.org/10.1016/j.v irusres.2013.01.017
- Izzati, U. Z., Inanaga, M., Hoa, N. T., Nueangphuet, P., Myint, O., Truong, Q. L., ... Yamaguchi, R. (2020).Pathological investigation and viral antigen distribution of emerging swine fever African in Vietnam. Transboundary and Emerging Diseases.
- Kolbasov, D., Titov, I., Tsybanov, S., Gogin, A., & Malogolovkin, A. (2018). African Swine Fever Virus, Siberia, Russia, 2017. Emerging infectious diseases, 24(4), 796-798. doi:10.3201/eid2404.171238

- Nga, B. T. T., Tran Anh Dao, B., Nguyen Thi, L., Osaki, M., Kawashima, K., Song, D., . . . Le, V. P. (2020). Clinical and Pathological Study of the First Outbreak Cases of African Swine Fever in Vietnam, 2019. *Frontiers in veterinary science*, 7(392). doi:10.3389/fvets.2020.00392
- OIE. (2019). African Swine Fever (Infection with African Swine Fever Virus). In Manual of Diagnostic Tests and Vaccines for Terrestrial Animals (8th ed., pp. 1-8).
- OIE. (2021). Follow-up report 3 African swine fever virus (Inf. with), Indonesia. Retrieved from https://wahis.oie.int/#/reportinfo?reportId=28198.
- Rodríguez-Bertos, A., Cadenas-Е., Fernández, Rebollada-Merino, A., Porras-González, N., Mayoral-Alegre, F. J., Barreno, L., . . . Sánchez-Vizcaíno, J. M. (2020).Clinical Course and Gross Pathological Findings in Wild Boar Infected with a Highly Virulent Strain of African Swine Fever Virus Genotype II. Pathogens (Basel, *Switzerland*), 9(9), 688. Retrieved from https://www.mdpi.com/2076-0817/9/9/688
- Salguero, F. J. (2020). Comparative Pathology and Pathogenesis of African Swine Fever Infection in Swine. *Frontiers*

*in veterinary science*, 7(282). doi:10.3389/fvets.2020.00282

Sánchez-Cordón, P. J., Jabbar, T., Chapman, D., Dixon, L. K., & Montoya, M. (2020). Absence of Long-Term Protection in Domestic Pigs Immunized Attenuated with African Swine Fever Virus Isolate OURT88/3 or Benin∆MGF Correlates with Increased Levels of Regulatory T Cells and Interleukin-10. Journal of Virology, 94(14), e00350-00320.

doi:10.1128/jvi.00350-20

- Sanchez-Cordon, P. J., Montoya, M., Reis, A. L., & Dixon, L. K. (2018). African swine fever: A re-emerging viral disease threatening the global pig industry. *Vet J*, 233, 41-48. doi:10.1016/j.tvjl.2017.12.02 5
- Sánchez-Cordón, P. J., Vidaña, B., Neimanis, A., Núñez, A., Wikström, E., & Gavier-Widén, D. (2021). Pathology of African swine fever. In L. Iacolina, M.-L. Penrith, S. Bellini, E. Chenais, F. Jori, M. Montoya, K. Ståhl, & D. Gavier-Widén (Eds.), *Understanding* and combatting African Swine Fever (pp. 87-139): Wageningen Academic Publishers.
- Sánchez-Vizcaíno, J. M., Mur, L., Bastos, A. D., & Penrith, M. L. (2015). New insights into the role of ticks in African

swine fever epidemiology. *Rev Sci Tech*, *34*(2), 503-511. doi:10.20506/rst.34.2.2375

Sánchez-Vizcaíno, J. M., Mur, L., Gomez-Villamandos, J. C., & Carrasco, L. (2015). An Update on the Epidemiology and Pathology of African Swine Fever. *Journal of Comparative Pathology*, *152*(1), 9-21. doi:https://doi.org/10.1016/j.j cpa.2014.09.003

Schulz, K., Staubach, C., & Blome, S. (2017). African and classical swine fever: similarities, differences and epidemiological consequences. Veterinary research, 48(1), 84-84. doi:10.1186/s13567-017-0490-x

- Urbano, A. C., Forth, J. H., Olesen, A., Dixon, L., Rasmussen, T. B., Cackett, G., ... Ferreira, F. (2021). African swine fever virus: cellular and molecular aspects. In L. Lacolina, M. L. Penrith, S. Bellini, E. Chenais, F. Jori, M. Montoya, K. Ståhl, & D. Gavier-Widén (Eds.), **Understanding** and combatting African Swine Fever (pp. 25-61). Wageningen: Wageningen Academic Publishers.
- Viltrop, A., Boinas, F., Depner, K., Jori, F., Kolbasov, D., Laddomada, A., . . . Chenais, E. (2021). African swine fever epidemiology, surveillance, and control. In L. Lacolina,

M. L. Penrith, S. Bellini, E. Chenais, F. Jori, M. Montoya, K. Ståhl, & D. Gavier-Widén (Eds.), *Understanding and combatting African Swine Fever* (pp. 229-261): Wageningen Academic Publishers.

- Yamada, М., Masujin, K., Kameyama, K.-I., Yamazoe, R., Kubo, T., Iwata, K., . . . Yamakawa, M. (2021). Experimental infection of pigs with different doses of the African swine fever virus Armenia 07 strain by intramuscular injection and direct contact. The Journal of veterinary medical science, 82(12), 1835-1845. doi:10.1292/jvms.20-0378
- Zhao, D., Liu, R., Zhang, X., Li, F., Wang, J., Zhang, J., ... Bu, Z. (2019). Replication and virulence in pigs of the first African swine fever virus isolated in China. *Emerging Microbes & Infections*, 8(1), 438-447. doi:10.1080/22221751.2019. 1590128
- Zhou, X., Li, N., Luo, Y., Liu, Y., Miao, F., Chen, T., . . . Hu, R. (2018). Emergence of African Swine Fever in China, 2018. *Transboundary and Emerging Diseases*, 65(6), 1482-1484. doi:https://doi.org/10.1111/tb ed.12989